

HELIANGOLIDES, KAURANES AND OTHER CONSTITUENTS OF *HELIANTHUS HETEROPHYLLUS*

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(Received 20 November 1985)

Key Word Index—*Helianthus heterophyllus*; Compositae; Heliantheae; heliangolides; sesquiterpene lactones; kauranes; diterpenes; norsesquiterpenes; 8-methoxyobliquin; coumarin.

Abstract—The chloroform extract of the aerial parts of *Helianthus heterophyllus* afforded the heliangolides 2',3'-dihydroleptocarpin and 2',3'-dihydroniveusin C and the new coumarin (–)-8-methoxyobliquin. Other constituents were various *ent*-kaurenoic acids, (+)-vomilifoliol, dehydromilifoliol, 9-hydroxy-4-megastigmen-3-one, 9,12,13-trihydroxy-octadeca-10(*E*),15(2*Z*)-dienoic and 9,12,13-trihydroxyoctadec-10(*E*)-enoic acid, 2,6-dimethoxybenzoquinone and vanillin.

INTRODUCTION

Since our most recent article on constituents of *Helianthus* species [1], a spate of reports on this subject, largely by Mabry and coworkers, has appeared [2–15]. We now describe the results of our study of *Helianthus heterophyllus* Nutt. A report from another source [16] has dealt with the flavonoids of *Helianthus* sect. *Divaricati*, ser. *Angustifolii* which includes *H. heterophyllus* [17].

RESULTS AND DISCUSSION

Like many other *Helianthus* species, *H. heterophyllus* furnished several *ent*-kaurenoic acids, specifically 1a–4a, which were identified in the form of their methyl esters. Small amounts of two heliangolides, 2',3'-dihydroleptocarpin (5a) [18] and 2',3'-dihydroniveusin C (6a), were also found. These were isolated and purified in the form of their pyrazoline adducts 7 and 8, analysis of whose ¹H NMR spectra (Table 1) permitted their identification. The paramagnetic shift ($\Delta\delta \sim 0.9$) of H-5 of 7, in comparison with H-5 of leptocarpin (5b) and its analogs, indicates that the stereochemistry of the pyrazoline moiety is as shown in the formula and predicted from the model. Likewise the H-5 signal of 8 is shifted downfield ($\Delta\delta \sim 0.3$) compared with H-5 of niveusin C (6b) and its analogs [19]. Lactone 6a appears to be new; its acetate 6c has been reported from a *Calea* species [20].

Another new compound was (–)-8-methoxyobliquin (9a), an isomer of 5-methoxyobliquin of unspecified rotation from *Helichrysum serpillifolium* [21]. That position C-5 on the coumarin nucleus was unsubstituted was evident from NOE experiments; irradiation at the frequency of H-4 at $\delta 7.57$ (Table 2) showed a clear NOE for the doublet at 6.28 (H-3, 32%) and the singlet at 6.79 (H-5, 20%). In the converse experiment, irradiation at the frequency of the H-5 signal enhanced the intensities of the H-4 signal (12%) and the signals of the vinylic protons at $\delta 4.45$ and 4.04, thus showing that the methoxy group was not on C-6 but on C-8, and that the five carbon moiety was attached to the coumarin nucleus in the manner of

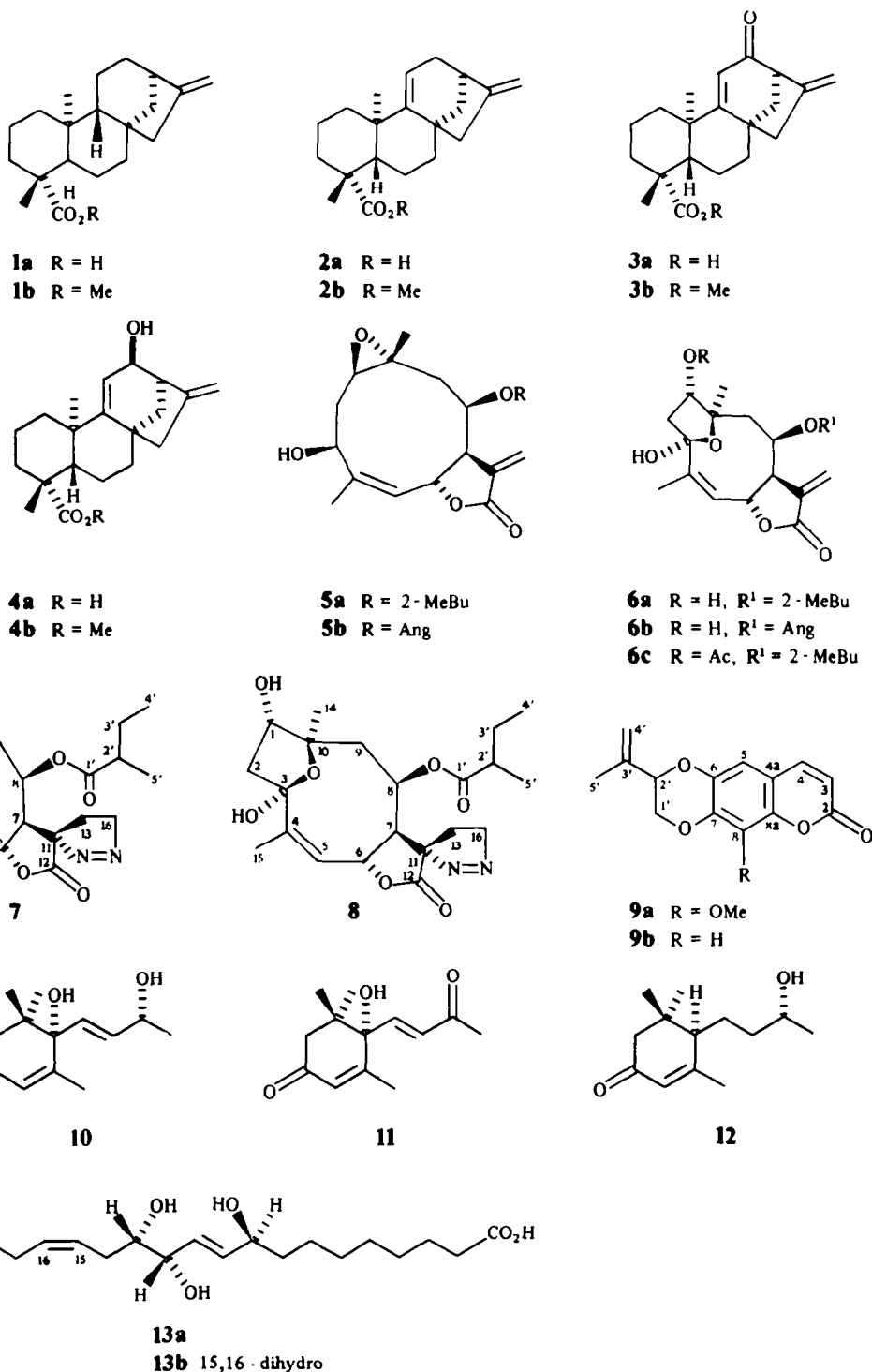
obliquin (9b) [22]. The rotation of 9a was negative, unlike that of (+)-obliquin whose absolute configuration is unknown.

The ¹³C NMR spectrum of 9a is listed in Table 3; assignments are based on chemical shifts and on analysis of the fully coupled ¹³C–¹H NMR spectrum with NOE enhancement. Thus the signal at $\delta 114.46$ was a sharp doublet with a C–H coupling constant of 173 Hz, while ¹J_{C–H} of the signal at $\delta 108.92$ which also exhibited a long

Table 1. ¹H NMR spectra of compounds 7 and 8 [270 MHz, CDCl₃; J (Hz) in parentheses]

H	7	8
1	2.86 <i>dd</i> (10, 5)	4.01 <i>dd</i> (8, 4.5)
2a	1.80 <i>ddd</i> (15, 10, 2.5)	2.56 <i>dd</i> (15, 4.5)
2b	2.50 <i>dt</i> (15, 5)	2.39 <i>d</i> (15)
3	4.54 <i>dd</i> (5, 2.5)	—
5	6.28 <i>dd</i> (12, 1)	5.97 <i>dd</i> (3, 1.5)
6	6.88 <i>br d</i> (12)	5.56 <i>ddd</i> (3, 2, 1.5)
7	2.45 <i>br s</i>	3.75 <i>dd</i> (4, 2)
8	5.03 <i>dd</i> (4.5, 2)	5.26 <i>ddd</i> (10, 6, 4)
9a	2.75 <i>dd</i> (15, 4.5)	2.07 <i>dd</i> (14, 10)
9b	1.20 <i>dd</i> (15, 2)	1.74 <i>dd</i> (14, 6)
13a	1.95 <i>m</i>	1.96 <i>m</i>
13b	1.65 <i>m</i>	1.40 <i>m</i>
14*	1.50 <i>s</i>	1.44 <i>s</i>
15*	1.89 <i>d</i> (1)	1.89 <i>t</i> (1.5)
16a	4.82 <i>m</i>	4.80 <i>ddd</i> (18, 10, 5)
16b	4.68 <i>m</i>	4.68 <i>ddd</i> (18, 10, 8)
2'	2.40 <i>tq</i> (7)	2.26 <i>tq</i> (7)
3'a	1.65 <i>m</i>	1.63 <i>m</i>
3'b	1.45 <i>m</i>	1.40 <i>m</i>
4*	0.90 <i>t</i> (7)	0.87 <i>t</i> (7)
5*	1.16 <i>d</i> (7)	1.07 <i>d</i> (7)
OH	2.18 <i>s</i>	2.95 <i>d</i> (8)
		3.26 <i>s</i>

*Intensity three protons.



range $^3J_{C-H}$ of ca 5 Hz was 163 Hz. Hence the signal at lower field was that of C-3, while the signal at higher field was that of C-5 [23], a conclusion which could be confirmed by selective decoupling. Among the quaternary carbons the signal at $\delta 112.69$ which showed only one $^3J_{C-H}$ of ~ 8 Hz could be assigned to C-4a because of its chemical shift and the complex signal at $\delta 136.02$ which

displayed several long range coupling constants could be identified as that of C-3'. Among the remaining three signals, that at $\delta 139.16$, which exhibited two $^3J_{C-H}$ s of about 8 Hz each, was therefore that of C-8a, the signal at $\delta 140.78$ with one long range coupling constant of ca 8 Hz was that of C-7 and the signal at $\delta 141.12$ with one long range coupling constant of 3.5 Hz ($^2J_{C-H}$) was that of C-6.

Table 2. ^1H NMR spectrum of compound **9a** [J (Hz) in parentheses]

H	δ
3	6.29 <i>d</i> (9.5)
4	7.57 <i>d</i> (9.5)
5	6.79 <i>s</i>
1'a	4.45 <i>dd</i> (11.5, 2.3)
1'b	4.04 <i>dd</i> (11.5, 8.2)
2'	4.54 <i>br dd</i> (8.2, 2.3)
4'a	5.19 <i>br s</i>
4'b	5.14 <i>br s</i>
5'	1.80 <i>br s</i>
OMe*	4.03 <i>s</i>

*Intensity three protons.

Table 3. ^{13}C NMR spectrum of compound **9a** (67.89 MHz, CDCl_3)

C	δ
2	160.50 <i>s</i>
3	114.46 <i>d</i>
4	143.39 <i>d</i>
4a	112.69 <i>s</i>
5	108.92 <i>d</i>
6	141.12 <i>s</i>
7	140.28 <i>s</i>
8	142.48 <i>s</i>
8a	139.16 <i>s</i>
1'	67.60 <i>t</i>
2'	75.63 <i>d</i>
3'	136.02 <i>s</i>
4'	114.88 <i>t</i>
5'	18.77 <i>q</i>
OMe	61.69 <i>q</i>

Other compounds identified in the *H. heterophyllus* extract were the bisnorsesquiterpenoids (+)-vomilifoliol (blumenol A, **10**) [24, 25], dehydrovomilifoliol (**11**) [26] and 9-hydroxy-4-megastigmen-3-one (**12**) [27], 2,6-dimethoxybenzoquinone, vanillin and two trihydroxy- C_{18} acids **13a**, **13b** first isolated by us from *Rudbeckia fulgida* [28] and later found in *Wedelia pinetorum* [29]. Two antifungal acids apparently identical with **13a**, **13b** have been isolated from a rice cultivar infected with rice blast disease [30] and are apparently present as such in resistant rice varieties [31]. Thus at least **13a** and **13b**, if not the other metabolites, appear to be part of the defense system of *H. heterophyllus*.

EXPERIMENTAL

Helianthus heterophyllus Nutt. was collected by Dr. John B. Nelson within the Apalachicola National Forest along Forest

Service Road # 344 1.5 miles So. of Hoxham, Leon County in October 1983. The dried aerial parts (2.27 kg) were extracted with CHCl_3 and worked up in the usual fashion [32]. The crude gum (3.2 g) was adsorbed on 6 g of silicic acid (Mallinckrodt 100 mesh) and chromatographed over 200 g of the same adsorbent packed in hexane, 125 ml fractions being collected as follows: 1–4 (hexane), 5–8 (hexane-EtOAc, 9:1), 9–12 (hexane-EtOAc, 4:1), 13–16 (hexane-EtOAc, 2:1), 17–20 (hexane-EtOAc, 1:1), 21–24 (hexane-EtOAc, 2:3), 25–28 (hexane-EtOAc, 1:4), 29–32 (EtOAc) and 33–36 (EtOAc-MeOH, 9:1).

Fr. 6 (0.04 g) was a mixture of **1a** and **2a** which was separated after methylation with CH_2N_2 and subsequent chromatography to give 7 mg of **1b** and 6 mg of **2b** identified by comparison with authentic material [33]. Fr. 9–10 (0.15 g) after purification by CC (C_6H_6 -EtOAc, 17:3) gave 35 mg of **9a** as a gum, $[\alpha]_{\text{D}}^{25} -29.5^\circ$ (CHCl_3 , c 0.420); MS m/z (rel. int.): 274 [M] $^+$ (100), 259 (1.4), 243 (20.4), 219 (19.11), 207 (3.4), 192 (10.4); ^1H and ^{13}C NMR spectra: Tables 2 and 3. [Calc. for $\text{C}_{15}\text{H}_{14}\text{O}_5$: M , 274.0841 Found: M , (MS) 274.0838].

Fr. 11–12 (0.25 g) were rechromatographed over 40 g of silica gel (70–230 mesh) packed in hexane. Elution with hexane-EtOAc (4:1) gave 5 mg of **9a**, 5 mg of vanillin and a mixture which was methylated with CH_2N_2 . Acetylation (Ac_2O -pyridine, overnight) and purification by TLC (hexane-EtOAc, 9:1, two developments) gave 4 mg of **3b** [34]. Fr. 13–14 (0.25 g) on initial purification by CC furnished impure **4a** which was methylated and then rechromatographed to give 10 mg of **4b** [35].

Reaction of fr. 15–16 (0.2 g) with CH_2N_2 and CC (CHCl_3) gave crude **7** and **8**. After rechromatography (hexane-EtOAc, 4:1), **7** was induced to crystallize (hexane-EtOAc, 9:1, -20°), mp 122° (dec); $[\alpha]_{\text{D}}^{25} -288^\circ$ (CHCl_3 , c 0.102); MS m/z (rel. int.): 406 [M] $^+$ (0.07), 378 (0.9), 360 (0.2), 276 (0.7), 258 (3.0), 233 (7.0), 232 (7.2), 177 (12.0), 163 (13.8), 149 (11.1), 135 (24.1), 95 (22.9), 85 (33.4), 57 (100); MS (CI) m/z (rel. int.): 407 [$\text{M} + 1$] $^+$ (0.45), 379 (100), 361 (2.2), 277 (33.3) and 259 (6.8). The ^1H NMR spectrum is listed in Table 1.

Crude **8** was purified by CC (hexane-EtOAc, 7:3) and crystallized from hexane-EtOAc (overnight, -20°), mp 149 – 150° ; $[\alpha]_{\text{D}}^{25} -253^\circ$ (CHCl_3 , c 0.072); IR $\nu_{\text{max}}^{\text{CHCl}_3}$ cm^{-1} : 3508, 1770, 1724 and 1600; MS m/z (rel. int.): 422 [M] $^+$ (0.4), 394 (0.6), 376 (0.7), 292 (4.7), 274 (3.6), 279 (20.8), 85 (21.5), 57 (100); MS (CI) m/z (rel. int.): 423 [$\text{M} + 1$] $^+$ (12.8), 395 (100), 377 (27.4), 293 (32.8) and 275 (46.8). The ^1H NMR spectrum is listed in Table 1.

Fr. 17–19 (0.28 g) on CC (hexane-EtOAc, 7:3) gave 5 mg of 2,6-dimethoxybenzoquinone and a mixture containing **11** and **12**. Separation of the mixture by prep. TLC (C_6H_6 -EtOAc, three developments) gave in the upper band 8 mg of **11** [25] and in the lower band 10 mg of **12** [26]. The ^1H NMR spectra of these substances coincided with those reported in the literature. Fr. 20–22 (0.2 g) containing one major component were purified by radial chromatography (hexane-EtOAc, 7:3) to give 60 mg of **10**, mp 110 – 112° (hexane-EtOAc), $[\alpha]_{\text{D}}^{25} +245^\circ$ (CDCl_3 ; c 0.08); ^1H NMR (270 MHz, CDCl_3): δ 5.90 ($J = 1$ Hz, H-4), 5.86 (*dd*, $J = 17.5$ Hz, H-8), 5.78 (*d*, $J = 17$ Hz, H-7), 4.41 (*dq*, $J = 6.5$, 5 Hz, H-9), 2.44 and 2.25 (AB system, $J = 17$ Hz, H-2a, b), 1.89 (*d*, $J = 1$ Hz, vinyl Me), 1.30 (*d*, $J = 6.5$ Hz, H-10), 1.08 (Me) and 1.01 (Me). Fr. 23–24 (0.2 g) on purification by CC (hexane-EtOAc, 3:7) gave 40 mg of a 1:1 mixture of **13a** and **13b** [27].

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